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Notes

pFJ269, A NEW PLASMID ISOLATED FROM A β -LACTAM ANTIBIOTIC PRODUCING STREPTOMYCETE

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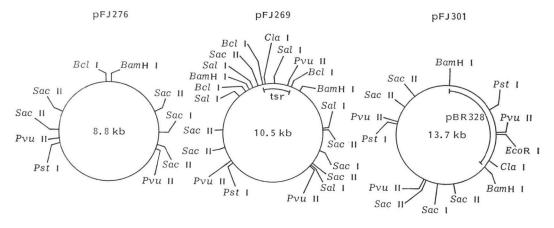
Streptomyces are Gram-positive, filamentous bacteria which have a complex life cycle that includes formation of substrate and aerial mycelia and spores. Streptomyces produce approximate-ly 70% of all naturally occurring antibiotics^{1,2)} and it is in the commercial production of antibiotics that this genus has been exploited. While the genetics and molecular biology of many Streptomyces are poorly understood, recent improvements in protoplast preparation, regeneration, fusion, and transformation^{3~6)} have facilitated the study of genetic organization, gene expression, and gene regulation in Streptomyces. Development of broad host range plasmid mediated gene cloning systems has also aided mole-

cular studies in *Streptomyces*. Although plasmid vectors currently available appear to have utility in many *Streptomyces* species^{7~9)}, restriction barriers between species and the possible instability of some plamids in a certain species could result in the need for different plasmid replicons. We report here the isolation and partial characterization of a new 8.8 kilobase plasmid from a β -lactam antibiotic producing streptomycete and its initial development into a gene cloning vehicle.

Total cellular DNA was isolated from Streptomyces strain A57284, an uncharacterized β lactam producer, by procedures cited in RICHARD-SON et al.8). Horizontal agarose gel electrophoresis revealed the presence of covalently closed circular DNA (cccDNA). The cccDNA was designated pFJ276. Although the strain from which pFJ276 was isolated produces a β -lactam antibiotic, no β -lactamase activity was detected in crude cell extracts. A restriction map of pFJ-276 (Fig. 1) was constructed following digestion with several restriction endonucleases in various combinations. A 1.7 kilobase BamH I fragment containing the Streptomyces azureus gene conferring resistance to thiostrepton, described by THOMPSON et al.⁶⁾, subcloned in our laboratory, was ligated to BamH I digested pFJ276. The

Fig. 1. Restriction endonuclease cleavage maps of pFJ276, pFJ269 and pFJ301.

pFJ276 was isolated from strain A57284. pFJ269 was constructed by cloning a 1.7 kb *Bam*H I fragment containing the *S. azureus* gene conferring resistance to thiostrepton into the unique *Bam*H I site of pFJ276. PFJ301 is a chimeric plasmid comprised of pBR328¹⁰⁾ and pFJ276, and can replicate and be amplified in *E. coli*. Many of the restriction sites of the pBR328 portion of pFJ301 have been omitted.



ligated DNA was used to transform Streptomyces ambofaciens protoplasts by the method cited in JONES et al.9). Transformants were selected for resistance to 50 µg/ml thiostrepton. The recombinant plasmid pFJ269 (Fig. 1) was found among the thiostrepton resistant transformants from which plasmid DNA was isolated. The copy number of pFJ269 in S. ambofaciens after 48 hours of growth in liquid culture was estimated by the method cited in JONES et al.9) to be about 40 copies per chromosome. Preliminary host range studies indicate that pFJ269 can function in S. ambofaciens and Streptomyces strain A49839, an uncharacterized β -lactam antibiotic producer that is different from Streptomyces strain A57284, the source of pFJ276. pFJ269 transforms S. ambofaciens at a frequency of about 10³ to 10⁴ transformants/ μ g. The initial transformation frequency of Streptomyces strain A49839 by pFJ269 was less than $50/\mu g$ because transformation procedures have not been optimized for this strain. The plasmid pFJ301 (Fig. 1) was constructed to serve as a convenient Escherichia coli source of pFJ276 by cloning pBR328 into the unique BamH I site of pFJ276.

pFJ269 is the first cloning vehicle with a replicon isolated from a β -lactam antibiotic producing streptomycete. Although it is at an early stage of development, pFJ269 is an example of an alternative plasmid replicon that could be useful in fundamental and applied studies in *Streptomyces*.

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